



The total synthesis of macrophelides A and E from carbohydrate precursors[†]

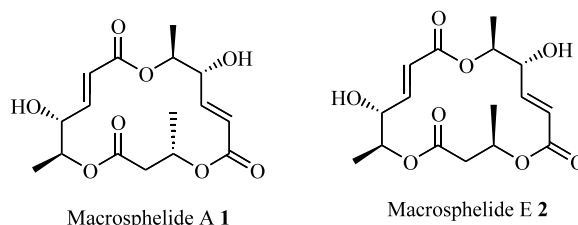
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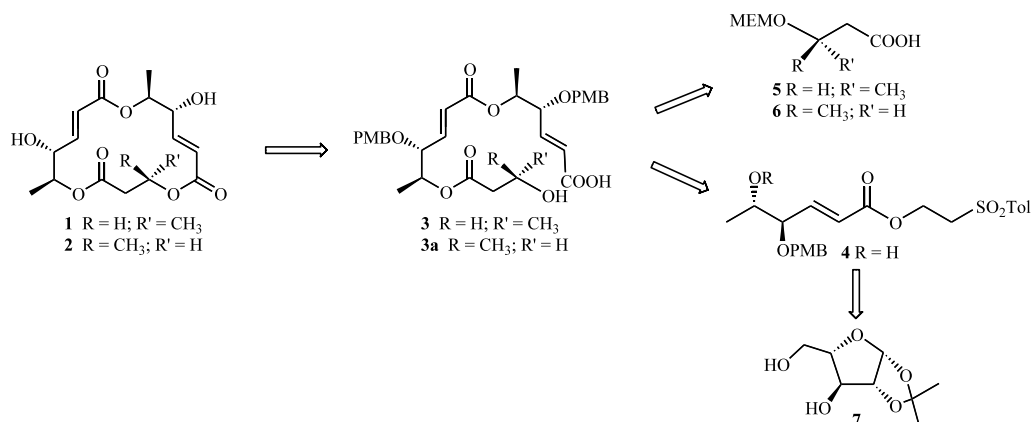
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Abstract—The total synthesis of macrolide antibiotics, macrophelide A and E has been achieved starting from carbohydrate precursors. © 2002 Elsevier Science Ltd. All rights reserved.

Macrophelides A–L are 16-membered macrolide antibiotics having three lactone bridges in the ring structure. Macrophelide A, isolated from the culture broth of *Microsphaeropsis* sp FO-5050, was found to inhibit strongly the adhesion of human leukemia HL-60 cells to an LPS-activated human-umbilical-vein endothelial cells (HUVETC) monolayer (IC₅₀, 3.5 μm) in a dose dependent fashion.¹ Macrophelide E, the C(3) stereoisomer of macrophelide A, was isolated from a strain of *Periconia byssoides* separated from the gastrointestinal tract of the sea hare *Aplysia kurodai*.² It was also found to inhibit the adhesion of human-leukemia HL-60 cells to human-umbilical-vein endothelial cells as potently as macrophelide A. Consequently, macrophelides have received much attention as valu-



able lead compounds for the development of new anti-cancer chemotherapeutic drugs, thus resulting in synthetic routes for generating these molecules and their analogues. Recently, the syntheses of macrophelides A 1^{3–5} and E 2⁵ has been reported. Herein, we report the first carbohydrate based total synthesis of macrophelides A 1 and E 2 from L (+)-arabinose.



Scheme 1. Retrosynthesis of macrophelide A 1 and E 2.

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The synthetic strategy presented here involved the preparation of *trans* (4*R*,5*S*)-5-hydroxy-4-*p*-methoxybenzyloxy-2-hexenoate **4**, condensation with commercially available 3*S*- or 3*R*-hydroxy butanoic acid unit **5** or **6**, respectively, and recoupling with one more unit of *trans* **4**, to give *seco* acids **3** and **3a**, respectively, which on Yamaguchi macrolactonisation afford the target molecules **1** and **2**. The retrosynthesis of macrophelides A and E is presented in Scheme 1.

Accordingly, 1,2-*O*-isopropylidene- β -L-arabinofuranose⁶ **7** was selectively tosylated (*p*-TsCl, pyridine, CH₂Cl₂) to give **8** in 75% yield, which on subsequent reduction with LiAlH₄ afforded **9** in 80% yield, [α]_D –12.6 (*c* 0.65, CHCl₃). Protection of hydroxy group in **9** with *p*-methoxybenzyl bromide (NaH, THF) afforded **10** (95%), which on hydrolysis of the 1,2-acetonide (cat. H₂SO₄ in 60% aq. AcOH) furnished **11** (75%). Oxidative cleavage of **11** (NaIO₄, MeOH–H₂O) and subsequent olefination of the unstable aldehyde **12** with (*p*-toluenesulfonyl)ethoxy carbonylmethylene triphenylphosphorane⁷ gave **13** in 75% yield. Finally, de-*O*-formylation of **13** with catalytic HCl (1,4-dioxane:water, 1:1) afforded the key intermediate **4** in 80% yield, [α]_D –39.1 (*c* 2.2, CHCl₃), in enantiopure form (Scheme 2).

Total synthesis of macrophelide A (**1**)

Condensation of alcohol **4** with acid **5** (Scheme 3) through the mixed anhydride prepared on reaction of **5** with 2,4,6-trichlorobenzoyl chloride (Et₃N, THF), in the presence of DMAP in toluene, afforded the ester **14** in 84% yield, [α]_D –39.3 (*c* 0.95, CHCl₃). Selective hydrolysis of the *p*-toluenesulfonyl ethyl group in **14** was effected with DBN⁷ (C₆H₆, rt) to furnish **15** in 81% yield. Esterification of carboxylic acid **15** with alcohol **4** using 2,4,6-trichlorobenzoyl chloride (Et₃N, THF) and DMAP in toluene gave **16** in 82% yield. Reaction of **16**

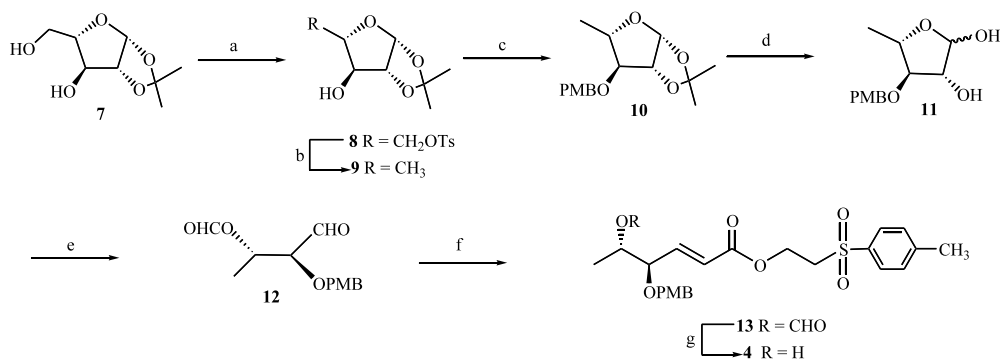
with TMSCl and NaI in CH₃CN (–20°C) afforded **17** in 77% yield, which on selective hydrolysis with DBN in C₆H₆ provided *seco* acid **3** in 79% yield. Finally, macrolactonisation of **3** under Yamaguchi conditions⁸ (2,4,6-trichlorobenzoyl chloride, Et₃N, DMAP, toluene) gave **18** in 67% yield, [α]_D –90.4 (*c* 0.215, CHCl₃), which on reaction with DDQ in CH₂Cl₂/H₂O gave **1** (90%), [α]_D +83.8 (*c* 0.10, MeOH); lit.¹⁵ [α]_D +84.1 (*c* 0.59, MeOH); lit.³ [α]_D +82 (*c* 0.10, MeOH); mp 139–141°C; lit.^{1b} mp 141–142°C.

Total synthesis of macrophelide E (**2**)

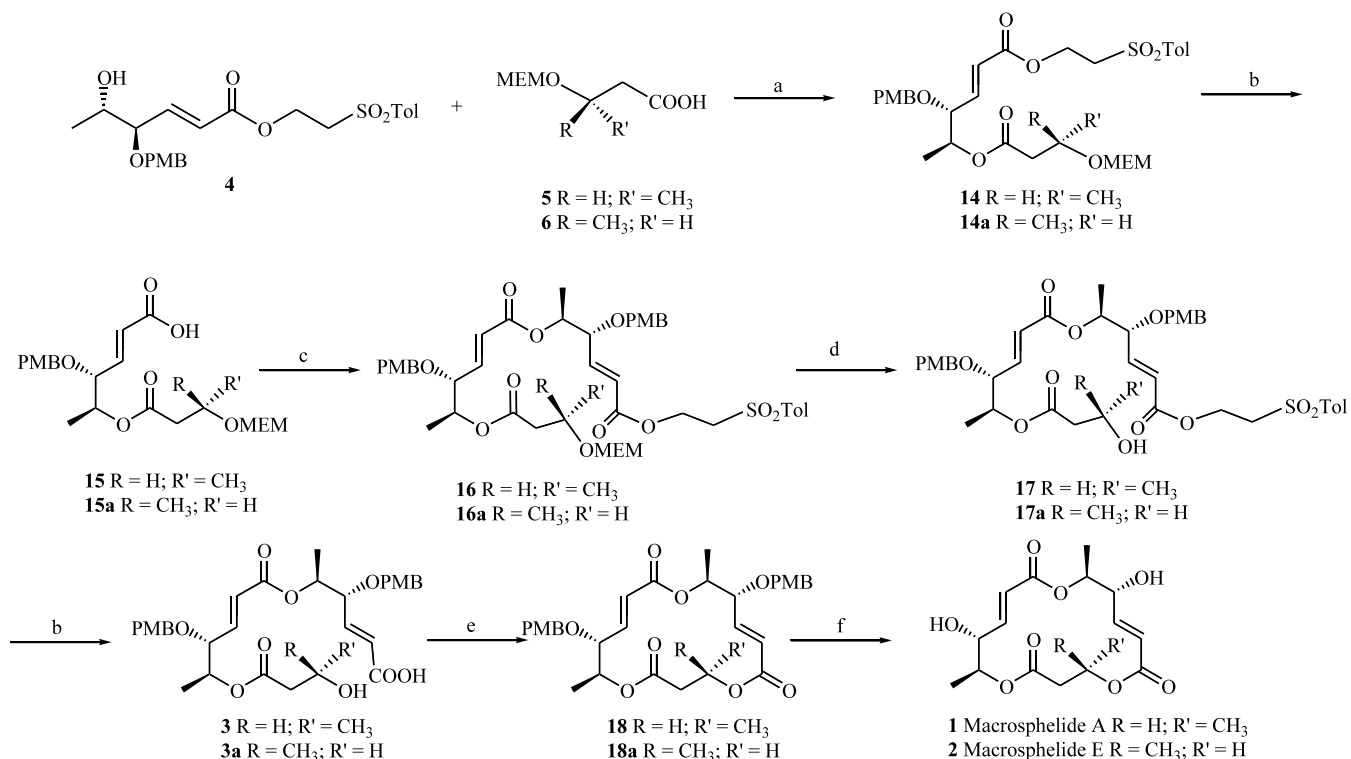
Similarly, esterification of acid **6** with alcohol **4** (Scheme 3) in the presence of 2,4,6-trichlorobenzoyl chloride (Et₃N, THF) and DMAP in toluene afforded the ester **14a** in 83% yield, [α]_D –13.13 (*c* 0.75, CHCl₃), which on selective hydrolysis with DBN in C₆H₆, furnished **15a** (80%). Esterification of carboxylic acid **15a** with alcohol **4** using 2,4,6-trichlorobenzoyl chloride (Et₃N, THF) and DMAP in toluene gave **16a** (84%), [α]_D –78.7 (*c* 0.25, CHCl₃). MEM deprotection of **16a** with TMSCl, NaI, in CH₃CN, afforded **17a** in 81% yield, which on selective hydrolysis with DBN in benzene gave **3a** (80%). Finally, **3a** under Yamaguchi macrolactonisation conditions (2,4,6-trichlorobenzoyl chloride, Et₃N, DMAP, toluene) gave **18a** in 70% yield. Deprotection of **18a** with DDQ in CH₂Cl₂/H₂O gave **2** in 93% yield, [α]_D +54.9 (*c* 0.28, EtOH); lit.^{2b} [α]_D +56.8 (*c* 0.46, EtOH); mp 104–106°C; lit.^{2b} mp 105–107°C. The target molecules **1** and **2** were fully characterised^{9,10} by ¹H, ¹³C NMR, FAB MS and IR spectra.

Acknowledgements

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Scheme 2. Reagents and conditions: (a) *p*-TsCl, pyridine, dry CH₂Cl₂, 0°C–rt, 14 h; (b) LiAlH₄, dry THF, 0°C–rt, 18 h; (c) NaH, PMBBr, dry THF, 0°C–rt, 4 h; (d) 60% aq. AcOH, cat. HCl, rt, 14 h; (e) NaIO₄, MeOH/H₂O (2:1), rt, 2 h; (f) Ph₃P=CHCOO(CH₂)₂SO₂Tol, 110°C, 1 h; (g) cat. HCl, dioxane/water (1:1), rt, 12 h.



Scheme 3. Synthesis of macrospheptide A **1** and E **2**. *Reagents and conditions:* (a) 2,4,6-trichlorobenzoyl chloride, Et₃N, THF, **4**, DMAP, toluene, rt, 12 h; (b) DBN, benzene, rt, 8 h; (c) 2,4,6-trichlorobenzoyl chloride, Et₃N, THF, **4**, DMAP, toluene, rt, 12 h; (d) TMSCl, NaI, CH₃CN, -20°C, 6 h; (e) 2,4,6-trichlorobenzoyl chloride, Et₃N, THF, DMAP, toluene, 90°C, 24 h; (f) DDQ, CH₂Cl₂/H₂O, rt.

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- Spectral data of macrospheptide-A **1**: IR (KBr): 3435, 1718 cm⁻¹; ¹H NMR (500 Hz, CDCl₃) δ 1.33 (d, 3H, *J*=6.6 Hz), 1.37 (d, 3H, *J*=6.3 Hz), 1.40 (d, 3H, *J*=6.6 Hz), 2.43 (d, 1H, *J*=6.9 Hz), 2.57 (dd, 1H, *J*=3.6, 15.7 Hz), 2.68 (dd, 1H, *J*=9.1, 15.7 Hz), 2.81 (d, 1H, *J*=7.4 Hz), 4.16 (br d, 1H, *J*=5.7 Hz), 4.23 (br d, 1H, *J*=5.2 Hz), 4.86 (dq, 1H, *J*=5.7, 6.3 Hz), 4.95 (dq, 1H, *J*=5.2, 6.3 Hz), 5.39 (m, 1H), 6.03 (dd, 1H, *J*=1.6, 9.3 Hz), 6.06 (dd, 1H, *J*=1.4, 9.0 Hz), 6.85 (dd, 1H, *J*=3.8, 15.6 Hz), 6.88 (dd, 1H, *J*=4.4, 15.6 Hz); ¹³C NMR (300 Hz, CDCl₃) δ 17.8, 18.0, 19.6, 40.9, 67.6, 73.1, 74.1, 74.8, 75.1, 122.3, 122.7, 144.9, 145.9, 164.4, 165.7, 170.1; FAB MS (*m/z*, %): 343 (M⁺+1, 2), 307 (28), 289 (17), 259 (8), 154 (57), 137 (100), 55 (58).
- Spectral data of macrospheptide-E **2**: IR (KBr): 3456, 1715, 1662 cm⁻¹; ¹H NMR (400 Hz, CDCl₃) δ 1.33 (d, 3H, *J*=6.1 Hz), 1.39 (d, 3H, *J*=6.1 Hz), 1.44 (d, 3H, *J*=6.7 Hz), 2.61 (dd, 1H, *J*=7.3, 15.8 Hz), 2.73 (dd, 1H, *J*=3.0, 15.8 Hz), 3.11 (d, 1H, *J*=6.7 Hz), 3.39 (d, 1H, *J*=7.9 Hz), 4.14–4.25 (m, 1H), 4.39 (br s, 1H), 4.99 (dq, 1H, *J*=4.2, 6.7 Hz), 5.13 (dq, 1H, *J*=2.0, 6.7 Hz), 5.34 (ddq, 1H, *J*=3.0, 6.7, 6.7 Hz), 6.08 (dd, 1H, *J*=1.2, 15.8 Hz), 6.12 (dd, 1H, *J*=1.2, 15.3 Hz), 6.81 (dd, 1H, *J*=4.8, 15.2 Hz), 7.03 (dd, 1H, *J*=4.3, 15.9 Hz); ¹³C NMR (300 Hz, CDCl₃) δ 17.3, 17.7, 19.5, 40.0, 66.6, 73.7, 75.1, 75.3, 75.9, 122.3, 122.9, 145.0, 145.3, 165.2, 166.6, 170.1; FAB MS (*m/z*, %): 343 (M⁺+1, 5), 307 (52), 289 (42), 259 (53), 221 (68), 207 (59), 176 (100).